

International Journal of Bio-pharmacology, Biotechnology and Allied Sciences

Review Article

www.ijbbas.in

THE INVERTASE- A REVIEW

Kumari N^1 , Sethy K^2

¹Department of Microbiology, Bundelkhand University, Jhansi, U.P.

²Centurion University Of Technology and Management, Bhubaneswar, Orissa.

*Corresponding Author: Neha kumari

Email ID: chiranshu.mrdls@gmail.com

Available online at: <u>www.ijbbas.in</u>.

Received 15th March. 2020; Revised 25th March. 2020; Accepted 5th April. 2020; Available online May. 2020

ABSTRACT

Invertase is an enzymes that catalyzes conversion of sucrose into glucose and fructose, is glycoprotein that works on pH 4.5 and stability at 50°c. it is broadly delivered in the biosphere in mostly microorganisms and plants. The present studies centered upon invertase together with source of production, isozymes, kinetics, media optimization, types of fermentation, purification methods and applications.

Key words: Bacterial Invertase, Beakers Yeast, Chromatography, Purification.

All living cells enclose convoluted globular protein named as enzymes, functions as biocatalyst expedites biochemical reactions in vivo and in vitro. Kuhan titled such biocatalyst as enzyme from Greek word, "enzumas" which point out brewing of bread by yeasts. The expedite nature of enzymes are accountable for the working. It cooperate in reaction without being depleted in the reaction, getting very high speed of product formation by decreasing the Gibbe's free energy (ΔG^{ϱ}) required to start the reaction.¹ Enzymes are very specific in nature due to this it can discriminate between chemicals with very much alike structure and expedite the reaction over different temperature range (0-110 °C) and in wide pH ranges (2-14). In industries, such properties with an enzyme being harmless and bio-degradable can results in large quantity and excellent stuff. Also an enzyme can be achieved from various microorganisms and that also enormous amount without applying any chemical resistant methods [1].

The industrial knowledge of enzymes rotates around malt and yeast where classical baking and leavening industries were immediately expanding. Much of the early progression of biochemistry was focused on yeast fermentations and mechanism for conversion of starch to sugar or cellulose to sugar **[2]**. one such types of enzyme of our attraction is "Invertase". This review centered on the catalytic nature, extraction methods and its application in today's globe.

The basic source of all living organism is carbohydrates. Monosaccharide like glucose and fructose are primarily used in metabolism [3]. Thus, invertase plays important role as it is sucrose degrading enzymes. ßfructofuranosidases (EC.3.2.1.26) or Invertase are special type of enzymes that drive the decompounds of sucrose into commixture of sucrose and glucose titled as inverted sugar [4]. decompounding of The b-Dfructuranosides (stachyose and raffinose) also accomplished by invertase whose products are beneficial in creation of bakery stuff and helps in brewing of sugarcane molasses [5]. Enzymatic action of invertase has been outlined chiefly in microorganism and plants.

Sources:

Plant Invertase:

There are three types of invertase namely, vacuolar, cytoplasmic and cell-wall, have been extracted from various species and characterized as biochemical level. Acid invertases have been extracted from various plants species. Soluble acidic invertase has been purified from many plant species, All soluble invertase inspected have a Km for sucrose in the mill molar range at a pH optimum between 4.5 and 5.5. In addition, all soluble invertase seem to be N- glycosylated proteins.

Table 1: Different plants sources were used to isolates invertase.

Plant species	Km in (mM)	References
Рарауа	7.7	[6]
Sugar cane	2.8	[7]
Oat	4.58	[8]
Maize	1.84	[9]
Potato	16	[10]

Bacterial sources:

There are various bacteria present in soil like Bacillus, and Bacillus cereus for invertase production. It is also found there are some gram negative bacteria those can discharge invertase enzyme relates to the order of actinomycetales such as Arthobacter species, Barevibacterium SP.

Table 2: Following bacteria were used forproduction of invertase enzyme.

Bacteria	Country	References
	&year	
Arthrobacter	2004 Japan	[11]
globiformis IFO		
3062		
Arthrobacter sp.	China 2009	[12]
10137		
Bacillus cereus	China 2007	[13]
TA-11		
Bacillus	2004 Cairo,	[14]
macerans	Egypt.	
Bifidobacterium	, Ireland 2005	[15]
infantis ATCC		
15697		
Brevibacterium	ND	[16]
divaricatum		
NRRL B		
Lactobacillus	ND	[17]
brevis Mm-6		
Thermotoga	ND	[18]
neapolitana		

Yeast sources:

Yeast is good sources of invertase enzyme production. Particularly Yeast has become very common source for invertase enzyme. Among the researchers many are utilizing yeast as rich source.

Table 3: Following yeast are used to produce

 invertase enzyme.

Yeast	Country	References
	year	
Candida		[19]
guilliermondii	México	
	2014	
Hansenula	2003 china	[20]
polymorpha		
Kluyveromyces	Pakistan	[21]
marxianus	2010	
Leucosporidium	2005	[22]
antarcticum	Honolulu	

Kinetics of Invertase:

Invertase show high catalytic activity over a wide range of pH (3.5-4.5) with the best pH of 4.5. The enzyme activity attains an optimum at 55 °C. The Michaelise Menten (Km) value for enzyme is 30 mM (approx.). This enzyme is glycoprotein stability shows at 50 °C. some important cataions like Hg²⁺ Ag⁺,Ca²⁺ and Cu²⁺ works as enzyme inhibitors **[23].** The fructose

analogue 2, 5-anhydro-D-mannitol shows Competitive inhibition instructing that the enzyme activity was forbidden by the furanose. AgNO₃ and HgCl₂ scrutinized as a total inhibiter of the enzyme while CuSO₄ and Cd(NO)₃ exerted a 76% and 73% inhibition, respectively **[23].**

Table 4: Some invertase inhibitors are listedhere.

Bacterial strain	Inhibitors	References
Arthrobacter	ND	[11]
globiformis		
IFO 3062		
Brevibacterium	Cu ^{2+,} SH	[16]
divaricatum		
NRRL B-2312		
Bacillus	Hg ²⁺ , Cd ²⁺	[25]
cereusTA-11	, Cu ²⁺	
Bifidobacterium	ND	[24]
breve		
UCC2003		
Bifidobacterium	Cu2+, Hg2 ⁺	[26]
infantis	рСМВ	
ATCC 15697		
Lactobacillus	Ca ^{2+,} Cu2 ^{+,} Cd ^{2+,}	[27]
reuteri CRL	Hg ^{2+,} β-ME,	
1100	DTT	
Thermotoga	2 mMCuSO ₄ ,	[28]
neapolitana	ZnSO ₄ , FeCl ₂ ,	
DSM 4359T	HgCl2,Urea,	

Review Article

Influence of pH on bacterial invertase activity:

Many research papers outlined the best pH for invertase enzyme activity is neutral pH with some exceptions .there were wide range of pH buffer applied to scrutinized the invertase activity. Different bacterial invertase works well on specific PH.

Microorg	pH and	Km	Referen
anism	temperature		ces
Brevibact	pH 6.8; 40 °C	0.19M	[16]
erium	more than		
divaricat	95% of the		
um NRRL	initial		
2312	activity was		
	at 30 °C and		
	less thar		
	15%at 50 °C		

pH 6.8; 37 °C

pH 7.0; 50 °C

[11]

[25]

Km

S

370

mΜ,

3.0

μmol

2.4Mm

,kO 127

Table 5: Effects of pH on invertase enzyme

		min-1	
Bifidobac	pH 6.0; 37 °C	Km-	[24]
terium		25mM	
breve		24	
UCC2003		µmol/	
		min/m	
		g	
Bifidobac	pl 4.3; pH	Vm/Km	[26]
terium	6.0;	= 0.65	
infantis	37 °C	and	
ATCC		0.025 ×	
15697		10-3	
		min-1	
		mg-1	
1	1		

Isozymes:

In yeast, there are two form of invertase that is extracellular invertase and intracellular Invertase [29]. Extracellular invertase is glycoprotein containing about 5% mannose, 50% carbohydrates, 3% glucosamine whereas intracellular invertase contains no carbohydrate. it has been scrutinized that in depressed yeasts invertase is extracellular and in well repressed state all invertase are intracellular. Bothe intracellular and extracellular enzymes are inhibited by iodine and mercaptoethanol reactivates it [30].

Arthroba

globiform

IFO 3062

Bacillus

cereus

TA-11

cter

is

In plats, many isoforms of invertase enzyme with different stands biocher idiosyncrasy. On the premise of solub isoelectric points, best pH and sub cel location, plant invertase can be categorized into three subgroups. These subgroups areacid soluble (vacuolar), alkaline soluble (cytoplasmic) and cell wall bound Invertase [31].

Invertase enzyme production from bacteria:

Optimized media for bacterial sources:

There is very less written matter focusing on media optimization of leavening condition for bacterial invertase production. Some studies have proclaimed Bacillus species an excellent source for invertase production and approved by food and drug administration. Bacteria are very fastidious in nature owe to this huge production of invertase enzyme can be enhanced by optimizing best production media. There are some production media has been used by researchers to enhance the production of invertase enzyme.

Table 6: Isolated bacterial strain, kind of enzyme, source and optimized production media

Bacteria	Kinds	Bacteria	Culture	Refer

mical		enzy
bility,		me
llular	Arthrob	Extrac

of

I source

	enzy		m	
	me			
Arthrob	Extrac		0.2%ye	[11]
acter	ellular		ast	
globifor			extract,	
mis			1%	
IFO			polype	
3062			ptone,	
			0.4%(N	
			H ₄) ₂ HP	
			O _{4,}	
			0.1%	
			MgS0 ₄	
			7H ₂ O,	
Arthrob	Extrac	Not	4%	[12]
acter sp.	ellular	identifie	corn	
10137		d	steep	
			powde	
			powde	
			powde r,	
			powde r, 0.13%	
			powde r, 0.13% MgSO4.	
			powde r, 0.13% MgSO ₄ . 7H ₂ O,	
			powde r, 0.13% MgSO ₄ . 7H ₂ O, yeast	
			powde r, 0.13% MgSO ₄ . 7H ₂ O, yeast extract,	
			powde r, 0.13% MgSO ₄ . 7H ₂ O, yeast extract, 0.4%(N	

ences

mediu

Review Article

Bacillus	Intrac	Soil	0.6%	[25]]				1%	
cereus	ellular		yeast						sucrose	
TA-11			extract,						,	
			0.1%			Bifidoba	Intrac	America	Synthe	[26]
			КН2РО			cterim	ellular	n Type	tic	
			4,0.1%			infantis		Culture	media	
			K ₂ HPO ₄			ATCC		Collecti	with	
			SY			15697		on	2%	
			broth					(ATCC)	fructos	
			with						е	
			1%			Brevibac	Intrac	Norther	1%	[16]
			sucrose			terim	ellular	n	urea, ,	
Bacillus	Intrac	National	3%sucr	[14]		divaricat		Regional	0.01%	
maceran	ellular	Researc	ose,0.5			um		Researc	biotin,	
S		h Centre	%			NRRL B-		h Lab	0.3%	
		(NRC)	pepton			2312		(NRRL)	KH ₂ PO ₄	
		Cairo,	e, 0.3%						7%	
		Egypt	yeast						sucrose	
			extract						,	
Bifidoba	Intrac	Infant	0.05%	[24]		Lactoba	Extrac	Milk of	10%	[17]
cterimbr	ellular	nursling	L-			cillus	ellular	breast	sucrose	
eve		stool	cystein			brevis			, 0.2%	
UCC200			e-HCl,			Mm-6			yeast	
3			Modifi						extract	
			ed			Lactoba	Intrac	CERELA	MRS	[32]
			Rogosa			cillus	ellular		broth	
			-			reuteri			added	
			Sharpe			CRL			with	
			mediu			1100			1%	
			m with						sucrose	

Review Article

Strepto	Extrac	Sugarca	Czapek	[33]
myces	ellular	ne field	-Dox	
sp.		soil	agar	
ALKC 8			(CDA)	
Thermot	Intrac	Deutsch	Resazu	[28]
oga	ellular	е	rin	
neapolit		Sammlu	0.0001	
ana		ng von	%, ,	
DSM		Mikroor	0.02%	
4359T		ganisme	MgCl ₂ .	
		n und	6H₂O,0	
		Zellkultu	$.1\% NH_4$	
		ren	Cl,	
		(Brauns	0.03%	
		chweig,	K ₂ HPO ₄	
		German	,0.03%	
		y)	KH_2PO_4	
			,	
			0.01%C	
			aCl ₂ .2H	
			₂ 0,0.1	
			%cystei	
			ne-HCl,	
			0.2%	
			yeast	
			extract,	
			0.01%	
			KCl,0.2	
			%trypt	
			one,0.5	

			%gluco	
			se,vita	
			min,1%	
			NaCl	
Zymomo	Extrac	CINVEST		[33]
nas	ellular	AV-IPN,	0.16%(
mobilis		Mexico	NH ₄)2S	
CDBB-B			O ₄ , 5%	
603			glucose	
			, 0.1%	
			MgSO ₄	
			· 7H₂O,	
			0.25	
			KH ₂ PO ₄	
			, 0.7%	
			bacto-	
			yeast	
			extract	

Favorable factors for fermentative condition:

Bacterial invertase production can be enhanced through the fermentation by optimizing the best factors those affects the fermentation process.

As compiled in the table, the production of bacterial invertase is positively affected by best incubation period (12-72h), optimum pH (4.5-9.7) and wide range of temperature (25-50).

Vol.1 (2), 192-209, May (2020)

Affects of agitation rate and inoculation level were not mentioned in most of the literature,

however 120 -250 rpm helps the growth of aerobic bacteria.

 Table 7: Optimized fermentation conditions for invertase production

Microorganis	incubatio	T°C	рН	Agitation	Inoculum	C source	N source	Referen
m	n			rate rpm	size			ce
Zymomonas	ND	30	4.9	No	ND	5%glucose	7%bacto-yeast	[11]
mobilis				agitation			extract,0.16%	
CDBB-B							(NH ₄) ₂ SO ₄	
603								
Streptomyces	24	37	5.0	ND	3 discs	1%	NaNO ₃ + yeast	[32]
sp.					(9 mm)	sucrose	extract	
ALKC 8								
Arthrobacter	36	25	7.0	200	ND	ND	1%polypepton	[11]
globiformis							e,0.2%yeast	
IFO							extract,0.4%	
3062							(NH ₄) ₂ HPO ₄	
Bacillus	36	50	9.5	100	ND	1%	0.6%yeast	[25]
cereus						sucrose	extract	
TA-11								
Bifidobacteri	16	37	6.8	ND	5%	2%fructos	Semi synthetic	[26]
um						е	medium	
infantis								
Lactobacillus	72	30	8.0	ND	2%	10%	2% malt	[17]
brevis Mm-6					(1.2 × 106	sucrose	extract,	
					CFU ml-1)		2% peptone	
Arthrobacter	20	30	ND	250	5%	4%	4% corn steep	[12]
sp.				(further		sucrose	powder, 0.4%	
10137				increased			(NH4) ₂ HPO ₄	
				by 20%)				

Influence of agitation rates on invertase production by p.expansum after 48 hr in culture media containing molasses as carbon source, yeast extract as nitrogen source, at 35°c, initial pH 5 a was inspected by using various agitation rate at 50,100, 250, and 300 rv/min. Maximum invertase production was found at 150 rv/min.

Enzyme purification:

Enzyme purification is process in which contaminates are removed to inspect the nature, function and structure of enzyme. Enzyme activity is enhanced by good purification strategies.

Purification methods	Bacterial strain	Purification	Yield %	Specific activity	Reference
SephacrylS-100; HPLC,Q-	Thermotoga	469.5	25.8	51,833.5	[28]
Sepharose F.F	neapolitana				
	DSM 4359T				
Gel filtration (Sephadex G-	Bacillus cereus	15.37	26.6	207.5	[25]
75), Ammonium sulfate	TA-11				
precipitation (40–80%);					
DEAE chromatography (A-					
50);					
Gel filtration (Sephadex	Lactobacillus	31.2	16.9	9.2	[27]
G-);Ionexchange	reuteri CRL				
chromatography (DEAE-	1100				
Sephacel), Ammonium					
sulfate precipitation					
(80%);					
Gel filtration (Sephadex G-	Bifidobacteriu	202	20.2	101	[34]
100),Chromatography on	т				
DEAE-Sepharose CL-6B,	adolescentis G1				
Toyopearl					
HW-65F Bio-Gel P-I00;					

Table 8: Enzyme purification approaches for bacterial invertase

DEAE-cellulose	Brevibacterium	345	8.2	345	[16]
chromatography;	divaricatum				
Ammonium sulfate precipitation	NRRL B-2312				
(48–70%);					

Production of invertase by submerged fermentation:

Currently, various microorganisms are desirable for production of invertase rather than plants or animals because of less toxic compounds are produced during fermentation process **[35]**.

Now days mostly submerged state fermentation process are used in invertase production compared to solid state fermentation [36].

On huge scale, submerged state fermentation is a classical fermentation process for production of invertase. It is profitable technology with maximum yielding per reactor volume and can be freely inflict to culture handling and downstream processing. In this fermentation production, the tabbed microorganism is cultured in sealed vessels that encompass oxygen and nutrient broth [**37**]. Invertase producing microorganisms utilizes nutrients for production of invertase and discharges it into the fermentation medium.

Differently, solid state fermentation manages microorganisms on solid base instead to liquid medium. This strategy arranges an environment that is very much alike to microbe's natural environment and appropriate for the growth of microorganism that need less moisture. Solid state fermentation is not adopted in immense production of invertase due to its restrains including deadlock to standardize the operations. The rising attentions on pollution and to examine cost effect have expanded the curiosity on application of waste materials for the production of Invertase [38].

Various waste materials and fermentative approaches have been out lined for invertase production as shown in table-

Table 9: Different waste materials andfermentation methods.

Substrate	Microorganism	Maximum production condition	Production	References
Agro-industrial wastes	A. niger	pH6.5 at 25°C for six days	15.9±2.44 u/g	[39]
Fruits peel	A. niger	pH5.0 at 30ºC for four days	16.25±0.60 μM	[40]
Molasses	Fusarium solani	pH5.0 at 30°C for four days	9.90 U/mL	[41]
Agriculture- based by- products	A. flavus	pH6.5 at 40°C for two days	7.41 U/mL	[42]

Submerged fermentation

Solid state fermentation

Substrate	microorganism	Production condition	yield	References
peels of fruit	A. niger	Four days at 30c ,pH 5	51 U/mL	[43]
Residue of red carrot	S. cerevisiae	72 hr at 30c	272.5 U/g	[43]
Residue of sugercane	S. cerevisiae	Three days at 40c ph 5	430 U/mg	[36]
Wheat bran	A. niger	Three days at 30c ph 5	194.71 U/g	[45]

It is difficult to find out the kinds of waste materials or strategies of fermentation compatible for invertase production since these researches are conducted in pilot scale using bioreactors. Thus the application of much alike approach in coming days will give more important information

Application of invertase:

Now day's enzymes are used in industrial actions because of their gentle performing condition like pH and temperature. In industries, Invertase has been employed in good application like food beverages, pharmaceutical and biosensor.

In beverages:

In beverages industries, invertase has been broadly used in manufacturing of fructose or invert syrup. Invertase is employed as and preferably to the classical acid hydrolysis approaches which might discharge unwanted by- products. Because of hygroscopic nature of invert sugar it is widely used in production of soft candies, jams, chocolates [36].

Pharmaceutical industry:

Differently, invertases are crucial enzyme in pharmaceutical industries for drug formulation. On the other hand, importance of invertase in pharmaceutical is enhanced by analysis of invertase with transfructosylating activities which generate FOS. FOS is an oligosaccharides made up of tiny fructose chains that is less sweeter than sucrose, less in calories, preparing them excellent for diabetic patients **[46]**.

Biosensor:

Invertase can help as biosensor for identification of sucrose in a simple and fast way rather than traditional methods such as electrophoresis and chromatography.

Bagal et al. (2015) stated the fiction of nano – gold cluster mediated by invertase, incorporate on onion membranes. This reported hypothesis can be employed as sucrose biosensors based on fluorescence [47]. Appropriate to more than one cancer is not an impossible achievement.

CONCLUSION

Invertase enzyme available in wide range of organisms encompasses immense commercial importance, especially in pharmaceutical and food industries. Different studies have earlier determined a full description of this enzyme but modern investigations which centered on sources of invertase that helps in isolation of invertase from different sources and might have exclusive trait such as transfructosylating action that will enhance the commercial value of this enzyme.

Furthermore, studies on isozymes of invertase and kinetics of invertase might be useful in creation artificial enzymes in future. Through the media optimization and type of fermentations and purification processes, yielding of invertase production can be enhanced by following compatible parameter.

REFERENCES

[1] Maciel, M. J. M., & Ribeiro, H. C. T. (2010). Industrial and biotechnological applications of ligninolytic enzymes of the basidiomycota: A review. *Electronic Journal of Biotechnology*, 13(6), 14-15.

[2] Abstract for Invert Sugar Production Line –Enzyme Company.

[3] Fotopoulos, V. (2005). Plant invertases: structure, function and regulation of a diverse enzyme family.

[4] Benattouche, Z., Raho, G. B., Bouhadi, D.,
& Sahnouni, F. (2014). Characterization of partially purified extracellular thermostable invertase by Streptococcus sp. isolated from dates. *Bulletin of Environmental, Pharmacology and Life Science*, 5(9), 6-72.

[5] Aziz, S., Jalal, F., Nawaz, M., Niaz, B., Ali Shah, F., Hafeez-ur-Rahman Memon, M., ... & Rajoka, M. I. (2011). Hyperproduction and thermal characterization of a novel invertase from a double mutant derivative of Kluyveromyces marxianus. *Food Technology and Biotechnology*, *49*(4), 465-473.

[6] CHAN JR, H. T., & KWOK, S. C. (1976). Isolation and characterization of a β -fructofuranosidase from papaya. *Journal of Food Science*, *41*(2), 320-323.

[7] Del Rosario, E. J., & Santisopasri, V. (1977). Characterization and inhibition of invertases in sugar cane juice. *Phytochemistry*, *16*(4), 443-445.

[8] Pressey, R., & Avants, J. K. (1980). Invertases in oat seedlings: separation, properties, and changes in activities in seedling segments. *Plant physiology*, *65*(1), 136-140.

[9] Doehlert, D. C., & Felker, F. C. (1987). Characterization and distribution of invertase activity in developing maize (Zea mays) kernels. *Physiologia Plantarum*, *70*(1), 51-57.

[10] Bracho, G. E., & Whitaker, J. R. (1990). Purification and partial characterization of potato (Solanum tuberosum) invertase and its endogenous proteinaceous inhibitor. *Plant physiology*, *92*(2), 386-394. [11] Win, T. T., Isono, N., Kusnadi, Y., Watanabe, K., Obae, K., Ito, H., & Matsui, H. (2004). Enzymatic synthesis of two novel nonreducing oligosaccharides using transfructosylation activity with β fructofuranosidase from Arthrobacter globiformis. *Biotechnology letters*, *26*(6), 499-503.

[12] Xu, Z. W., Li, Y. Q., Wang, Y. H., Yang, B., & Ning, Z. X. (2009). Production of β fructofuranosidase by Arthrobacter sp. and its application in the modification of stevioside and rebaudioside A. *Food technology and biotechnology*, 47(2), 137-143.

[13] Yoon, M. H., Choi, W. Y., Kwon, S. J., Yi, S. H., Lee, D. H., & Lee, J. S. (2007). Purification and properties of intracellular invertase from alkalophilic and thermophilic Bacillus cereus TA-11. *Journal of Applied Biological Chemistry*, *50*(4), 196-201.

[14] Ahmed, S. A. (2008). Invertase production by Bacillus macerans immobilized on calcium alginate beads. *J Appl Sci Res*, *4*(12), 1777-1781.

[15] Vásquez-Bahena, J. M., Vega-Estrada, J., Santiago-Hernández, J. A., Ortega-López, J., Flores-Cotera, L. B., Montes-Horcasitas, M. C., & Hidalgo-Lara, M. E. (2006). Expression and improved production of the soluble extracellular invertase from Zymomonas mobilis in Escherichia coli. *Enzyme and microbial technology*, *40*(1), 61-66.

[16] Yamamoto, K., Kitamoto, Y., Ohata, N., Isshiki, S., & Ichikawa, Y. (1986). Purification and properties of invertase from a glutamateproducing bacterium. *Journal of Fermentation Technology*, *64*(4), 285-291.

[17] Awad, G. E., Amer, H., El-Gammal, E. W., Helmy, W. A., Esawy, M. A., & Elnashar, M. M. (2013). Production optimization of invertase by Lactobacillus brevis Mm-6 and its immobilization on alginate beads. *Carbohydrate polymers*, *93*(2), 740-746.

[18] Dipasquale, L., Gambacorta, A., Siciliano, R. A., Mazzeo, M. F., & Lama, L. (2009). Purification and biochemical characterization of a native invertase from the hydrogenproducing Thermotoganeapolitana (DSM 4359). *Extremophiles*, *13*(2), 345. [19] Vásquez-Bahena, J. M., Vega-Estrada, J., Santiago-Hernández, J. A., Ortega-López, J., Flores-Cotera, L. B., Montes-Horcasitas, M. C., & Hidalgo-Lara, M. E. (2006). Expression and improved production of the soluble extracellular invertase from Zymomonas mobilis in Escherichia coli. *Enzyme and microbial technology*, 40(1), 61-66.

[20] Plascencia-Espinosa, M., Santiago-Hernández, A., Pavón-Orozco, P., Vallejo-Becerra, V., Trejo-Estrada, S., Sosa-Peinado, A., ... & Hidalgo-Lara, M. E. (2014). Effect of deglycosylation on the properties of thermophilic invertase purified from the yeast Candida guilliermondii MpIIIa. *Process Biochemistry*, *49*(9), 1480-1487.

[21] Acosta, N., Beldarraín, A., Rodríguez, L., & Alonso, Y. (2000). Characterization of recombinant invertase expressed in methylotrophic yeasts. *Biotechnology and applied biochemistry*, *32*(3), 179-187.

[22] Aziz, S., Jalal, F., Nawaz, M., Niaz, B., Ali Shah, F., Hafeez-ur-Rahman Memon, M., ... & Rajoka, M. I. (2011). Hyperproduction and thermal characterization of a novel invertase from a double mutant derivative of Kluyveromyces marxianus. *Food Technology and Biotechnology*, *49*(4), 465-473. **[23]** Ali, S., & Haq, I. (2007). Kinetics of improved extracellular β -d-fructofuranosidase fructohydrolase production by a derepressed Saccharomyces cerevisiae. *Letters in applied microbiology*, *45*(2), 160-167.

[24] Ryan, S. M., Fitzgerald, G. F., & van Sinderen, D. (2005). Transcriptional regulation and characterization of a novel β fructofuranosidase-encoding gene from Bifidobacterium breve UCC2003. *Appl. Environ. Microbiol.*, *71*(7), 3475-3482.

[25] Yoon, M. H., Choi, W. Y., Kwon, S. J., Yi, S. H., Lee, D. H., & Lee, J. S. (2007). Purification and properties of intracellular invertase from alkalophilic and thermophilic Bacillus cereus TA-11. *Journal of Applied Biological Chemistry*, *50*(4), 196-201.

[26] Warchol, M., Perrin, S., Grill, J. P., & Schneider, F. (2002). Characterization of a purified β -fructofuranosidase from Bifidobacterium infantis ATCC 15697. *Letters in Applied Microbiology*, *35*(6), 462-467.

[27] De Ginés, S. C., Maldonado, M. C., & De Valdez, G. F. (2000). Purification and characterization of invertase from Lactobacillus reuteri CRL 1100. *Current Microbiology*, *40*(3), 181-184. **[28]** Dipasquale, L., Gambacorta, A., Siciliano, R. A., Mazzeo, M. F., & Lama, L. (2009). Purification and biochemical characterization of a native invertase from the hydrogenproducing Thermotoganeapolitana (DSM 4359). *Extremophiles*, *13*(2), 345.

[29] Lahiri, S., Basu, A., Sengupta, S., Banerjee, S., Dutta, T., Soren, D., ... & Ghosh, A. K. (2012). Purification and characterization of a trehalase–invertase enzyme with dual activity from Candida utilis. *Archives of biochemistry and biophysics*, *522*(2), 90-99.

[30] Workman, W. E., & Day, D. F. (1983). Purification and properties of the β -fructofuranosidase from Kluyveromyces fragilis. *FEBS letters*, *160*(1-2), 16-20.

[31] Kim, D., Lee, G., Chang, M., Park, J., Chung, Y., Lee, S., & Lee, T. K. (2011). Purification and biochemical characterization of insoluble acid invertase (INAC-INV) from pea seedlings. *Journal of agricultural and food chemistry*, *59*(20), 11228-11233.

[32] Kaur, N., & Sharma, A. D. (2005). Production, optimization and characterization of extracellular invertase by an actinomycete strain. **[33]** Vásquez-Bahena, J. M., Vega-Estrada, J., Santiago-Hernández, J. A., Ortega-López, J., Flores-Cotera, L. B., Montes-Horcasitas, M. C., & Hidalgo-Lara, M. E. (2006). Expression and improved production of the soluble extracellular invertase from Zymomonas mobilis in Escherichia coli. *Enzyme and microbial technology*, *40*(1), 61-66.

[34] Muramatsu, K., Onodera, S., Kikuchi, M., & Shiomi, N. (1993). Purification and some properties of β -fructofuranosidase from Bifidobacterium adolescentis G1. *Bioscience, biotechnology, and biochemistry*, *57*(10), 1681-1685.

[35] Benattouche, Z., Raho, G. B., Bouhadi, D., & Sahnouni, F. (2014). Characterization of partially purified extracellular thermostable invertase by Streptococcus sp. isolated from dates. *Bulletin of Environmental, Pharmacology and Life Science, 5*(9), 6-72.

[36] Kumar, R., & Kesavapillai, B. (2012). Stimulation of extracellular invertase production from spent yeast when sugarcane pressmud used as substrate through solid state fermentation. *SpringerPlus*, 1(1), 81.

[37] Pang, W. C., Ramli, A. N. M., & Johari, N. D. (2019). Structural Properties, Production, and Commercialisation of Invertase. *Sains Malaysiana*, *48*(3), 523-531..

[38] Nadeem, H., Rashid, M. H., Siddique, M. H., Azeem, F., Muzammil, S., Javed, M. R., ... & Riaz, M. (2015). Microbial invertases: a review on kinetics, thermodynamics, physiochemical properties. *Process Biochemistry*, *50*(8), 1202-1210.

[39] Al-Hagar, O. E., Ahmed, A. S., & Hassan, I. A. (2015). Invertase production by irradiated Aspergillus niger OSH5 using agricultural wastes as carbon source. *Microbiology Research Journal International*, 135-146.

[40 Mehta, K., & Duhan, J. S. (2014). Production of invertase from Aspergillus niger using fruit peel waste as a substrate. *Int. J. Pharm. Bio. Sci*, *5*(2), 353-360.

[41] Bhatti, H. N., Asgher, M., Abbas, A., Nawaz, R., & Sheikh, M. A. (2006). Studies on kinetics and thermostability of a novel acid invertase from Fusarium solani. *Journal of agricultural and food chemistry*, *54*(13), 4617-4623.

[42] Ahmed, K., Valeem, E. E., & Mahmood, T. (2015). Optimal cultural conditions for industrial enzyme production by using shaken

flask technique of submerged fermentation. *FUUAST Journal of Biology*, 5(1), 21-26.

[43] Raju, A. I. C. H., Pulipati, K., & Jetti, A. (2016). Production of Invertase by Aspergillus niger Under Solid State Fermentation Using Orange Fruit Peel as Substrate. *Adv Crop Sci Tech*, *4*(247), 2.

[44] Rashad, M. M., & Nooman, M. U. (2009). Production, purification and characterization of extracellular invertase from Saccharomyses Cerevisiae NRRL Y-12632 by solid-state fermentation of red carrot residue. *Australian Journal of Basic and Applied Sciences*, *3*(3), 1910-1919.

[45] Esawy, M. A., Kansoh, A. L., Kheiralla, Z. H., Ahmed, H. E., Kahil, T. A. K., & El-Hameed, E. K. (2014). Production and immobilization of halophilic invertase produced from honey isolate Aspergillus niger EM77 (KF774181). *International Journal of Biotechnology for Wellness Industries*, *3*(2), 36-45.

[46] Dominguez, A. L., Rodrigues, L. R., Lima, N. M., & Teixeira, J. A. (2014). An overview of the recent developments on fructooligosaccharide production and applications. *Food and bioprocess technology*, *7*(2), 324-337.